Detection of *Taylorella equigenitalis* and *Taylorella asinigenitalis* in horses in Croatia as a result of small scale survey

Zrinka Štritof1*, Josipa Habuš1, Vesna Mojčec Perko1, Melita Majhut2, Nika Brkljača Bottegaro3, Matko Perharić1, Suzana Hadina1, Zoran Milas1, and Nenad Turk1

1Department of Microbiology and Infectious Diseases with Clinic, Faculty of Veterinary Medicine, University of Zagreb, Zagreb, Croatia
2Faculty of Veterinary Medicine, University of Zagreb, Zagreb, Croatia, student
3Clinic of Surgery, Orthopedics and Ophthalmology, Faculty of Veterinary Medicine, University of Zagreb, Zagreb, Croatia


**ABSTRACT**

Contagious equine metritis (CEM) is a highly contagious venereal disease caused by the bacterium *Taylorella equigenitalis*. In Croatia, horses are almost never tested for this disease unless it is specifically requested. To date, no positive bacteriology results for *T. equigenitalis* have ever been obtained in Croatia. During 2014 and 2015, 12 animals were screened for *T. equigenitalis* using both bacteriology and a polymerase chain reaction (PCR) test. No positive cultures were obtained, however, samples from three animals tested positive on PCR. Sequencing of the PCR products revealed that one animal was positive for *T. equigenitalis* and two animals for *T. asinigenitalis*. These results represent the first evidence of *T. equigenitalis* and *T. asinigenitalis* infections in horses in Croatia. The high prevalence of *T. equigenitalis* and *T. asinigenitalis* positive horses (n = 3) in the population tested (n = 12) strongly suggests that the overall prevalence of horses positive for *Taylorella* spp. in Croatia is high, and that the equine population, especially those animals used for breeding, should be regularly tested for this bacterium. In addition, these results show the low sensitivity of using culture alone and that additional molecular methods should be utilised to identify positive cases of *Taylorella* spp.

**Key words:** horse, *Taylorella equigenitalis*, *Taylorella asinigenitalis*, Croatia

*Corresponding author:*

Zrinka Štritof, Department of Microbiology and Infectious Diseases with Clinic, Faculty of Veterinary Medicine, University of Zagreb, Heinzeloova 55, 10 000 Zagreb, Croatia, Phone: +385 1 2390 200; Fax: +385 1 2390 207; E-mail: zstritof@vef.hr
Introduction

Contagious equine metritis (CEM) is a highly contagious venereal disease caused by the bacterium *Taylorella equigenitalis*, a capnophilic, non-motile, small Gram-negative rod or pleomorphic coccobacillus (PLATT et al., 1977; RICKETTS et al., 1977). The disease was first described in the United Kingdom in 1977 (CROWHURST, 1977; PLATT and ATHERTON, 1977; TIMONEY et al., 1977). Subsequently, CEM has been found in many other countries in Europe and throughout the world (SWERCZEK, T. W., 1978; BLOBEL et al., 1979; HAZARD et al., 1979; KAMADA et al., 1981; MAY et al., 2012; RICKETTS et al., 2012). In stallions, *T. equigenitalis* is merely commensal, permanently colonizing specific sites on the external genitalia, without causing clinical signs (POWELL, 1981). Stallions therefore serve as a primary reservoir of this bacterium. In mares, *T. equigenitalis* infection may be subclinical or may manifest as endometritis, cervicitis and/or vaginitis, which results in temporary infertility (TIMONEY and POWELL, 1988). Mares may also develop a carrier state which can last from several months to years (HEATH and TIMONEY, 2008). Transmission of *T. equigenitalis* occurs mechanically during mating or via contamination upon semen collection. The rate of transmission via natural service exceeds that from either chilled or extended semen (SCHULMAN et al., 2013). A similar bacterium from the same genus, *T. asinigenitalis*, was first isolated from a donkey in 1997 (KATZ et al., 2000; JANG et al., 2001) and thereafter from a horse in 2004 (BAVERUD et al., 2006). The clinical relevance of *T. asinigenitalis* is currently unclear and is probably negligible (JANG et al., 2001; MEADE et al., 2010). However, due to their similar growth requirements, morphology and biochemical characteristics, it is important to differentiate *T. asinigenitalis* from *T. equigenitalis*. This can be achieved by using molecular methods, with species specific primers, preferably qPCR, which also increases test sensitivity and significantly shortens turnaround time (WAKELEY et al., 2006; MAY et al., 2016).

In Croatia, there is currently very little pre-breeding testing for *T. equigenitalis* in either mares or stallions, even as a part of any investigation into poor conception rates in mares. The objectives of this study were to screen a small population of equids for *T. equigenitalis*, to determine the prevalence of this disease in Croatia and to raise awareness of its presence in the general equine population, and more specifically, of the possible consequences for mare fertility.

Materials and methods

Samples. In 2014 and 2015, a total of twelve animals were tested (ten stallions and two mares). Samples from three stallions were submitted to the laboratory for pre-breeding CEM testing requested by the owner, while samples from the other nine animals (seven stallions and two mares) were collected at the Faculty of Veterinary Medicine, University of Zagreb, Croatia, from stallions who were there for routine castration of...
stallions and during pre-breeding examinations of mares. In stallions, swabs (Nuova Aptaca SRL, Canelli, Italy) were obtained from four predilection sites: the distal urethra, the fossa glandis, the urethral sinus and the preputial folds. In mares, swabs were taken from two predilection sites: the clitoral fossa and the clitoral sinuses. Paediatric swabs (Copan Innovation, Brescia, Italy) were used for sampling of the clitoral sinus. Only one set of swabs was obtained from each animal. Immediately after collection, the swabs were transported in Amies transport medium with charcoal (Nuova Aptaca SRL, Canelli, Italy), to the laboratory under temperature-controlled conditions, and plated within 24 hours of collection.

**Culture.** Culture was performed according to the OIE Manual of Diagnostic Tests and Vaccines for Terrestrial Animals (OIE, 2012). All swabs were inoculated onto two types of media: 1) chocolate agar with horse blood supplemented with antibiotics (clindamycin, trimetoprim sulfonamid, amfotericin B) as previously described (TIMONEY et al., 1982) and 2) chocolate agar with horse blood and the addition of amphotericin B (10 mg/mL). Inoculated plates were incubated in microaerophilic conditions (with addition of 7% CO₂) for 10 days, and examined on the 3rd, 7th and 10th days of incubation. Used plates were simultaneously tested with control strains of *T. equigenitalis* and *T. asinigenitalis*.

**Polymerase chain reaction (PCR) and sequence analysis.** After plating, each swab was washed thoroughly with 500 µL phosphate buffer saline (PBS), from which DNA was extracted from the suspension using a commercial kit (Qiagen, Netherlands) according to manufacturer’s instructions. Primers targeting the 16S rRNA gene were used for PCR as previously described (BLEUMINK-PLUYM et al., 1994). The cycling conditions were as follows: denaturation: 30 s at 94 °C; primer annealing: 15 s at 55 °C; and extension: 1 min at 72 °C. PCR products were electrophoresed in 2% agarose gel and visualized with Gel Doc System (Bio-Rad, CA, USA). As the primers described by BLEUMINK-PLUYM et al., 1994 do not discriminate *T. equigenitalis* from *T. asinigenitalis*, products obtained by PCR were sequenced and analyzed using Molecular Evolutionary Genetics Analysis software (MEGA 6.06). Sequence analysis of the 16S rRNA genes was used to differentiate *T. equigenitalis* from *T. asinigenitalis*, and to identify the *Taylorella* spp. (BAVERUD et al., 2006).

**Results**

All cultures were either negative or were contaminated before the end of the incubation period of ten days. In 3/12 (25%) animals, a PCR product of 585 bp was obtained from at least one predilection site. The primers used, as described by BLEUMINK-PLUYM et al., 1994, amplify a product of 585 bp in *Taylorella* spp. positive samples. For practical purposes, the positive animals were assigned the letters A, B and C. Animal A was a stallion whose urethral fossa was positive, but the distal urethra and penile sheath were
negative. Animal B was a stallion whose urethral fossa and distal urethra tested positive, while the penile sheath was negative. Animal C was a mare whose clitoral fossa and clitoral sinuses tested positive. These results are summarized in Table 1. All bands correlated with the positive controls, as well as with the adequate size of the molecular marker (Fig. 1). Sequence analysis of the PCR products revealed that of the three positive animals, one was positive for *T. equigenitalis* (animal A) and two for *T. asinigenitalis* (animals B and C).

<table>
<thead>
<tr>
<th>Animal</th>
<th>Clitoral fossa</th>
<th>Clitoral sinuses</th>
<th>Urethral sinus</th>
<th>Urethral fossa</th>
<th>Distal urethra</th>
</tr>
</thead>
<tbody>
<tr>
<td>A (stallion)</td>
<td>Not applicable</td>
<td>Not applicable</td>
<td>Negative</td>
<td>Positive</td>
<td>Negative</td>
</tr>
<tr>
<td>B (stallion)</td>
<td>Not applicable</td>
<td>Not applicable</td>
<td>Negative</td>
<td>Positive</td>
<td>Positive</td>
</tr>
<tr>
<td>C (mare)</td>
<td>Positive</td>
<td>Positive</td>
<td>Not applicable</td>
<td>Not applicable</td>
<td>Not applicable</td>
</tr>
</tbody>
</table>

**Discussion**

Interestingly, the animals that tested positive for *Taylorella spp.* had rarely been bred and none of them had had any contact with donkeys. Animal A was a warmblood stallion that had been bred only once, several years before. Animal B was a coldblood stallion that had been bred three times, and animal C was a warmblood mare whose reproductive history was unknown to the owner, but she had not been bred for at least three years prior to sampling. In all three animals it was impossible to determine the source of infection. Nine in-contact animals from the same premises as the positive animals all tested negative for *Taylorella spp.*

In Croatia, equine reproductive problems, both in terms of low conception rates and abortions, are rarely adequately addressed or their cause investigated. Despite the fact that all expenses incurred during the investigation of an abortion are covered by the Government, horse owners are still reluctant to report an abortion to a veterinarian. This is probably due to the fact that in cases where an infectious cause is determined, the owners are required to carry out mandatory disease control measures, including transport restrictions. In most cases where an abortion is reported, only samples for serology from the mare are submitted. Therefore, viral causes of reproductive disorders such as Equine Herpes Virus and Equine Arteritis Virus are readily diagnosed. After abortion, mares are also tested for leptospirosis, which forms an integral part of an abortion diagnostics panel (TURK et al., 2013). Nevertheless, evidence of other bacterial causes of reproductive problems in horses is scarce. The findings from our study strongly suggest
that *T. equigenitalis* could be a cause of the low conception rates seen in Croatian horses (PRVANOVIĆ et al., 2008).

The fact that we detected *T. equigenitalis* in such a small sample size is concerning. It indicates that the overall prevalence of *T. equigenitalis* infection in horses in Croatia may be high and requires further investigation of a larger sample size.

*T. asinigenitalis* was detected in 2/12 (16%) animals. Although the clinical relevance of *T. asinigenitalis* is currently unknown and probably negligible (JANG et al., 2001), awareness of its presence in the Croatian equine population is relevant in order to distinguish it from *T. equigenitalis* during diagnostic testing.

**Conclusion**

In conclusion, considering the control measures for CEM instituted in most European countries and the ever increasing transportation of horses, both nationally and internationally, it would be beneficial to have an insight into the epidemiological status of this disease in Croatia. Our findings highlight the need for the implementation of surveillance and control measures for CEM in Croatia. As a minimum, mandatory testing of all licensed stallions in Croatia should be introduced in order to estimate disease prevalence, minimise reproductive loss and to prevent the further spread of this disease.

---

**Acknowledgements**

The authors would like to thank Irena Zdovc from the Institute of Microbiology and Parasitology, Veterinary Faculty, University of Ljubljana, Slovenia, for her highly appreciated advice and suggestions and also Petra Pavečič for her excellent technical assistance.

**References**


Z. Štritof et al.: Detection of *Taylorella* spp. in horses in Croatia


Z. Štritof et al.: Detection of *Taylorella* spp. in horses in Croatia


Received: 27 June 2016
Accepted: 20 January 2017


**SAŽETAK**


**Ključne riječi:** konj, *Taylorella equigenitalis*, *Taylorella asinigenitalis*, Hrvatska